

^1H NMR (90 MHz, CDCl_3): δ 1.60 (s, 6H), 1.67 (s, 3H) and 1.75 (s, 3H) due to $4 \times \text{Me}$ groups; 1.90–2.20 (m, 8H) due to $4 \times$ allylic CH_2 groups; 3.45 (br d, $J = 7$ Hz, 2H: H-1'); 5.19 (m, 2H: H-6' and H-10'); 5.45 (br t, $J = 7$ Hz, 1H: H-2'); 6.85 (br s, 1H: H-2); 7.0–7.5 (complex m, 3H: H-5, H-6 and H-7); 7.6–7.7 (m, 1H: H-4) and 7.95 (br s, 1H: N-H); CI-MS (CH_4) m/z (rel. int.): 321 [M] $^+$ (16); 184 [$\text{M} - \text{C}_{10}\text{H}_{17}$] $^+$ (10.5); 158 [$184 - \text{CN}$] $^+$ (5.3) and 130 [$\text{M} - \text{C}_{14}\text{H}_{23}$] $^+$ (100) and ^{13}C NMR resonances as shown in Table 1.

Acknowledgements—We are grateful to Mr L. B. Mwasumbi, Department of Botany, University of Dar es Salaam and Dr B. Verdcourt, Kew Royal Botanic Gardens, England, for their efforts in the identification of the plant and to Prof. B. Zwanenburg and Mr L. Thijs, University of Nijmegen, The Netherlands, for the spectra.

REFERENCES

1. Achenbach, H. and Raffelsberger, B. (1979) *Tetrahedron Letters* 2571.
2. Achenbach, H., Renner, C. and Addae-Mensah, I. (1984) *Heterocycles* 22, 2501.
3. Benesova, V., Samek, Z., Herout, V. and Sorm, F. (1969) *Collect. Czech. Chem. Commun.* 34, 1807.
4. Plieninger, H. and Sirowei, H. (1971) *Chem. Ber.* 104, 2027.
5. Ishi, H. and Muzakani, Y. (1975) *Tetrahedron* 31, 933.
6. Jerram, W. A., McInnes, A. G., Maas, W. S. G., Smith, D. G., Taylor, A. and Walter, J. A. (1975) *Can. J. Chem.* 53, 727.
7. Gomez, F., Quijano, L., Calderon, J. S., and Rios, T. (1980) *Phytochemistry* 19, 2202.

Table 1. Tentative assignment of ^{13}C NMR (15.08 MHz, CDCl_3) spectral data for 3-farnesylindole (1)

Chemical shift (δ)	Assignment
136.1	C-8
135.2	C-7'
134.6	C-3'
130.7	C-11'
127.1	C-9
124.0	C-10'
123.8	C-2
122.5	C-6
121.4	C-5
120.7	C-2'
118.7 (br)	C-4 and C-6
110.4 (br)	C-3 and C-7
38.7 (br)	C-4' and C-8'
25.6 (br)	C-12'
25.6 (br), 24.5 and 22.7	C-9', C-5' and C-1'
16.5	C-3'Me
14.9 (br)	C-7'Me and C-11'Me

br = Broad signal.

8. Crombie, L., King, R. W., and Whiting, D. A. (1975) *J. Chem. Soc. Perkin Trans I*, 913.
9. Johnson, L. F., and Jankowski, W. C. (eds) (1972) "Carbon-13 NMR spectra". Wiley-Interscience, New York.

Phytochemistry, Vol. 26, No. 8, pp. 2403–2405, 1987.
Printed in Great Britain.

0031-9422/87 \$3.00+0.00
© 1987 Pergamon Journals Ltd.

TWO GUAIANOLIDES FROM *CENTAUREA COLLINA*

ISABEL FERNÁNDEZ, BEGOÑA GARCÍA, FRANCESC J. GRANCHA and JOSÉ R. PEDRO*

Department of Organic Chemistry, Faculty of Chemistry, University of Valencia, Burjassot, Valencia, Spain

(Revised received 8 February 1987)

Key Word Index—*Centaurea collina*; Compositae; sesquiterpene lactones; guaianolides.

Abstract—Two guaianolides isolated from the aerial parts of *Centaurea collina* were identified as 3β -hydroxy- 8α -epoxymethylacriloloxo-4(15),10(14),11(13)-trien-($1\alpha\text{H}$),($5\alpha\text{H}$)-guaian-6,12-olide and its 11 β ,13-dihydro derivative by spectroscopic methods.

INTRODUCTION

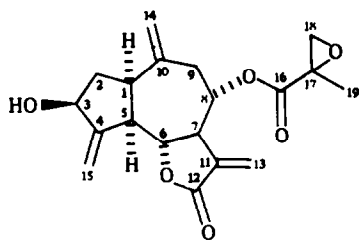
Many sesquiterpene lactones have been reported from the large genus *Centaurea* (Compositae, Cynareae) with approximately 700 species [1]. In the present paper we

report the isolation and structure elucidation of two guaianolides from *C. collina*, one of them being new. In previous work [2–4] several flavonoids were reported from this plant.

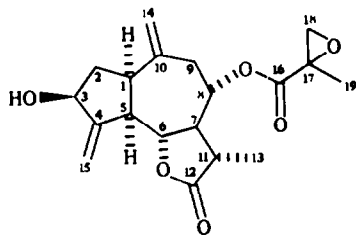
RESULTS AND DISCUSSION

From the methanolic extract of the aerial parts of *C. collina*, two sesquiterpene lactones were isolated.

*To whom correspondence should be addressed.



1



2

Compound (1) was non-crystalline; its IR spectrum showed bands characteristic of a γ -lactone (1750 cm^{-1}), a hydroxyl group ($3500\text{--}3200\text{ cm}^{-1}$) and a double bond (1650 cm^{-1}). The high resolution mass spectrum showed a $[M]^+$ at m/z 346.1421 corresponding to $C_{19}H_{22}O_6$, with fragments at m/z 262 and 244 which suggested the presence of a C_4 ester side chain. The $^1\text{H NMR}$ data (Table 1) established that the side chain was an epoxymethylacrylic acid residue (singlet at δ 1.34 and two doublets typical of an AB system at δ 3.57 and 3.78 ($J = 11.7\text{ Hz}$). The exocyclic methylene conjugated with a carbonyl lactone was easily recognizable in the two one-proton-doublets at δ 5.91 and 6.14 ($J = 3.0\text{ Hz}$). Two pairs of broad singlets (δ 5.30, 5.40 and δ 4.81, 5.07) were attributed to the hydrogens of an exocyclic methylene group attached to C-4 and C-10, respectively. The lactonic proton (H-6) appeared as a triplet at δ 4.22, the coupling constant of which ($J = 9.0\text{ Hz}$) indicated its *trans*-diaxial disposition to the hydrogens at C-5 and C-7. The H-8 proton gave a doublet triplet at δ 5.05 ($J = 9.0$ and 4.0 Hz) the coupling of which indicated the α -orientation for the ester group. The chemical shifts (δ 4.49) and coupling constant (t , $J = 7.5\text{ Hz}$) for H-3 indicated a β -orientation for the 3-hydroxyl group [5]. These signals, as were all the remaining signals of the $^1\text{H NMR}$ spectrum, were assigned by spin decoupling. According to the above data, the structure (1) was assigned to this compound which has been recently isolated from *C. solstitialis* [6].

Compound (2) ($C_{19}H_{24}O_6$, $[M]^+$ 348.1609) was non-crystalline. In the IR spectrum, absorption for a hydroxyl group was observed at $3550\text{--}3200\text{ cm}^{-1}$, for a γ -lactone at 1770 cm^{-1} , for an ester at 1740 cm^{-1} and for a double bond at 1640 cm^{-1} . As in 1, the presence of an epoxymethylacrylic ester was revealed by ions at m/z 264 and 246 in the mass spectrum and the signals at δ 1.35(s), 3.57 and 3.78 (AB system, $J = 11.2\text{ Hz}$) in the $^1\text{H NMR}$ spectrum (Table 1). The $[M]^+$ at m/z 348 is two mass units greater

Table 1. $^1\text{H NMR}$ spectral data for compounds 1 and 2

	1	2
H-1	2.91 <i>q</i>	2.91 <i>q</i>
H-2	1.68 <i>dt</i>	1.70 <i>ddd</i>
H-2'	2.11 <i>ddd</i>	2.20 <i>m</i>
H-3	4.49 <i>t</i>	4.53 <i>tt</i>
H-5	2.72 <i>t</i>	2.77 <i>t</i>
H-6	4.22 <i>t</i>	4.14 <i>t</i>
H-7	3.06 <i>tt</i>	2.24 <i>q</i>
H-8	5.05 <i>dt</i>	5.00 <i>dt</i>
H-9	2.30 <i>dd</i>	2.23 <i>dd</i>
H-9'	2.60 <i>dd</i>	2.70 <i>dd</i>
H-11	—	2.54 <i>dq</i>
H-13	5.91 <i>d</i>	1.30 <i>d</i>
H-13'	6.14 <i>d</i>	—
H-14	4.81 <i>s</i>	4.96 <i>s</i>
H-14'	5.07 <i>s</i>	5.12 <i>s</i>
H-15	5.30 <i>s</i>	5.30 <i>t</i>
H-15'	5.40 <i>s</i>	5.40 <i>t</i>
H-18	3.57 <i>d</i>	3.57 <i>d</i>
H-18'	3.78 <i>d</i>	3.78 <i>d</i>
H-19	1.34 <i>s</i>	1.35 <i>s</i>

Coupling constants (Hz): 1, $J_{1,2} = J_{1,2'}$ = $J_{1,5} = 9$; $J_{2,2'} = 13$; $J_{2,3} = J_{2,3'} = 7.5$; $J_{5,6} = J_{6,7} = J_{7,8} = 9$; $J_{8,9} = J_{8,9'} = 4$; $J_{9,9'} = 15$; $J_{7,13} = J_{7,13'} = 3$; $J_{18,18'} = 11.7$. 2, $J_{1,2} = J_{1,2'} = J_{1,5} = 9$; $J_{2,2'} = 13.5$; $J_{2,3} = J_{2,3'} = 7$; $J_{5,6} = J_{6,7} = J_{7,8} = J_{7,11} = 9$; $J_{8,9} = J_{8,9'} = 4$; $J_{9,9'} = 14$; $J_{11,13} = 7$; $J_{3,15} = J_{5,15} = 1.8$; $J_{18,18'} = 11.2$.

than that of 1 and this difference is maintained in the most significant fragment ions (m/z 264 and 246). In the $^1\text{H NMR}$ spectrum of 2, the signal corresponding to the conjugated exocyclic methylene does not appear, although a doublet is observed at δ 1.30 ($J_{11,13} = 7.0\text{ Hz}$) which may be attributed to a C-11 methyl group. The α -orientation of this methyl group is inferred from the coupling constant ($J_{7,11} = 9.0\text{ Hz}$) [7, 8]. The signals corresponding to H-14, H-15, H-3, H-6, H-8 (Table 1) and spin decoupling experiments allowed us to assign the structure (2) for this compound.

Further characterization of the guaianolides (1) and (2) was obtained by $^{13}\text{C NMR}$ spectroscopy. The data (Table 2) were interpreted on the basis of previous assignments [9, 10] and distortionless enhancement by polarization transfer (DEPT) experiments. Although some resonances were not assigned without ambiguity, it is clear that (1) and (2) possess the same relative stereochemistry C-1,3,5-8.

EXPERIMENTAL

MS were recorded at 70 eV. NMR spectra were measured at 200 MHz for ^1H and 50 MHz for ^{13}C . *C. collina* L. was collected at the Enguera-Ayora road (Valencia). A voucher specimen is deposited at the Botanic Department Herbarium, Faculty of Biological Sciences, University of Valencia. Aerial parts (3.6 kg) were extd exhaustively firstly with hexane and afterwards with

Table 2. ^{13}C NMR spectral data for compounds 1 and 2

	1	2
C-1	45.3 ^a	44.5
C-2	38.3 ^b	38.7 ^b
C-3	73.2	73.3
C-4	151.8	152.4
C-5	51.5	52.2 ^a
C-6	78.1 ^c	78.7 ^c
C-7	46.9 ^a	51.0 ^a
C-8	75.2 ^c	77.7 ^c
C-9	35.6 ^b	39.0 ^b
C-10	141.6	141.8
C-11	136.8	41.3
C-12	169.5	178.4
C-13	123.6	15.2
C-14	118.1	117.5
C-15	113.8	112.8
C-16	174.8	175.0
C-17	76.0	75.9
C-18	68.1	68.0
C-19	21.6	21.8

^{a,b,c} Chemical shifts denoted by the same letter in each column may be interchanged.

MeOH. The MeOH extract was reduced *in vacuo* to ca 1 l diluted with H_2O (2l) and re-extd with Et_2O . Evapn of solvent yielded 56 g of crude syrup which was chromatographed on silica gel. Elution of the column with mixtures of increasing polarity (hexane- CH_2Cl_2 -EtOAc) and repeated CC of the fractions eluted with CH_2Cl_2 -EtOAc (9:11) afforded 1 (30 mg) and 2 (15 mg).

Compound (1). Non-crystalline. IR ν_{max} cm^{-1} : 3500–3200,

2930, 1750, 1650, 1460, 1270, 1140, 1070, 990 and 930. MS m/z (rel. int.): 346.1421 $[\text{M}]^+$ (0.6) ($\text{C}_{19}\text{H}_{22}\text{O}_6$ requires 346.1410) 262 (6.0) 244 (39.3) and 226 (15.6).

Compound (2). Non-crystalline. IR ν_{max} cm^{-1} : 3550–3200, 2930, 1770, 1740, 1640, 1460, 1380, 1190, 1140, 1060, 1010 and 910. MS m/z (rel. int.): 348.1609 $[\text{M}]^+$ (0.5) ($\text{C}_{19}\text{H}_{24}\text{O}_6$ requires 348.1509) 264 (4.6), 246 (33.6) and 228 (13.8).

Acknowledgements—Financial support by the Comisión Asesora de Investigación Científica y Técnica (CAICYT, Grant No. 559/84) is gratefully acknowledged. We thank Prof. Dr J. Mansanet, Department of Botany, Faculty of Biological Sciences, University of Valencia (Spain) for identification of plant material.

REFERENCES

1. Fischer, N. H., Oliver, E. J. and Fischer, H. D. (1979) *Fortschr. Chem. Org. Naturst* **38**, 47.
2. Kamanzi, K., Raynaud, J. and Voirin, B. (1982) *Pharmazie* **37**, 454.
3. Kamanzi, K., Raynaud, J. and Voirin, B. (1982) *Plant. Med. Phytother.* **16**, 30.
4. Kamanzi, K., Raynaud, J. and Voirin, B. (1982), *Pharmazie* **37**, 523.
5. Bohlmann, F. and Zdero C. (1982) *Phytochemistry* **21**, 647.
6. Jakupovic, J., Jia, Y., Pathak, V. P., Bohlmann, F. and King, R. M. (1986) *Planta Med.* 399.
7. Das, S., Baruah, R. N., Sharma, R. P., Baruah, J. N., Kulanthaivel, P. and Herz, W. (1983) *Phytochemistry* **22**, 1989.
8. Bohlmann, F., Sing, P., Jakupovic, J. and Huneck, S. (1985) *Planta Med.* 74.
9. Massiot, G., Morfaux, A., Le Men-Olivier, L., Bouquant, J., Madaci, A., Mahamoud, A., Chopova, M. and Aclinou, P. (1986) *Phytochemistry* **35**, 258.
10. González Collado, I., Macías, F. A., Massanet, G. M. and Rodríguez Luis, F. (1986) *Tetrahedron* **42**, 3611.